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## OPTIMIZATION OF ULTRASOUND-ASSISTED ALKALINE EXTRACTION OF PUMPKIN SEED MEAL PROTEIN ISOLATE BY RESPONSE SURFACE METHODOLOGY

*As an alternative to animal proteins, pumpkin seed meal protein isolate (PSMPI) can be applied in food processing as dietary supplements and functional foods. The conventional extraction process of protein is alkaline solution and acid precipitation, which consumes a long time. Ultrasound method is an environmentally friendly technique, which can be used to improve the efficiency of protein extraction. In the present study, a method of ultrasound-assisted alkaline extraction (UAAE) of PSMPI was established. Response surface methodology (RSM) was utilized to optimize the UAAE conditions (ultrasonic power; ultrasonic time and ratio of liquid to solid) by implementing a three-factor, three level Box–Behnken design. A quadratic polynomial equation was obtained by polynomial regression analysis of the experimental data and appropriate statistical methods. The interaction effects of the independent variables on the extraction yield of PSMPI were illustrated by response surfaces and counter plots. Besides, by solving the regression equation and analyzing the response surface contour plots, the determined optimal PSMPI extraction conditions were obtained, which was at ultrasonic power of 456 W, ultrasonic time of 22 min, and ratio of liquid to solid of 27 mL/g. The efficiency of UAAE on PSMPI was determined. The result showed that the experimental value (80.76 g/100g) of the yield of PSMPI was reasonably close to the predicted one (81.86 g/100g). This suggested that there was a high fit degree between experimental value and the predicted value from the regression model. Thus, it suggested that the equation of the response surface can be utilized effectively to predict the extraction condition of the PSMPI.*

**Key words:** Box-behnken, ultrasound-assisted alkaline extraction, pumpkin seed meal protein isolate, optimize model.

**Introduction.** Protein is not only a necessary nutrient for the development and maintenance of human health, but also plays an important role in the physicochemical properties and structural basis of food [1, p. 30]. At present, animal protein is still an important source of dietary protein to most people. According to the report, the world demand for animal-derived proteins is supposed to double by 2050 [2, p. 94]. However, the production of animal proteins gives a great pressure on the environment. Besides, the popularity of vegetarianism has increased greatly over the past few years on account of growing concerns about the environment, animal welfare, and health. Thus, the utilization of plant protein sources (like oilseeds, cereals and legumes) as replacers for animal proteins is thought to be a good way to meet the need for food proteins.

**An analysis of the latest research and publications.** The most commonly used method to extract

proteins from plant protein sources is alkaline solution and acid precipitation [3, p. 1843]. However, due to the long extraction process, it will lead to protein denaturation. The reason is that the stability of secondary, tertiary, or quaternary structures changes, as well as the possible adverse secondary reactions and potential toxicity, which may lead to aggregation, solidification, reduced solubility, and loss of functional properties of proteins, such as gelation and emulsification [4, p. 703].

Enzymatic extraction is another widely used technique to isolate protein. The enzyme-assisted extraction of proteins could damage the cell walls integrity due to the degradation of the major components of plant cell walls and fiber, including cellulose, hemicellulose, and/or pectin caused by enzymatic action, as well as the improvement in protein solubility as a result of partial hydrolysis by different proteases. The increased degradation of

cell walls could enhance the protein release into the solvent. However, this method involves high energy consumption, harmful chemicals and high labor cost [5, p. 206].

Recently, as environmentally friendly techniques, the application of microwave-assisted and ultrasound-assisted extraction attracts considerable attention. The microwave power of MAE provides more uniform heating, producing a strong pressure on the plant cell walls, as a result, the desirable components could be broken and be dissolved in the solvent. The primary advantages of microwave are easy to handle, less solvent use, less waste generation, high extraction speed [6, p. 4667]. Recent studies have shown that some functional properties (water and nitrogen solubility index and foam stability) of proteins may decrease while some functional properties (fat absorption index, water absorption index, foam activity, emulsifying activity, and in vitro protein digestibility) will also be improved after microwave treatment [7, p. 543].

Ultrasound technology makes use of acoustic cavitation to disrupt plant cells and tissues and improve mass transfer during the extraction process. The application of ultrasound to assist extraction has the advantages of easy handling and safety, because it is carried out at atmospheric pressure and ambient temperature, uses an appropriate amount of solvent, is reusable and requires relatively short processing time [8, p. 50; 9, p. 813]. It was reported that ultrasound-assisted extraction significantly increased protein yield from rice bran, soy bean, perilla seed and pumpkin seed. However, high power and long time of sonication may reduce emulsification and foaming capability [2, p. 93]. Notably, according to the previous study, both of the ultrasound treatment and microwave treatment could decreasing the allergenicity of soybean protein isolate due to the alteration of secondary structure of proteins [10, p. 410; 11, p. 604].

Pumpkin seeds are a by-product in the utilization of pumpkin, which protein content ranges from 24.5–36% [12, 701]. The defatted pumpkin seed meal was obtained after cold-pressing oil extraction from pumpkin seeds and is considered to be a potential source of proteins for human consumption [13, p. 470]. The main constituents of pumpkin seed proteins are: albumin (water-soluble), globulin (salt-soluble), glutenin (alkali-soluble) and proline (alcohol-soluble) [14, p. 591]. It has been demonstrated that the essential amino acids content of globulin, glutelin and prolamin in pumpkin seed meet the minimum FAO/WHO/UNU needs

for preschool children, except threonine and lysin [15, p. 7715]. The pumpkin seed meal could be transferred to additional value products such as protein concentrate or isolate and applied in food processing as dietary supplements and functional foods.

The purpose and the objectives of the study.

The main goal of the investigation is – to optimize the ultrasound-assisted alkaline extraction (UAAE) conditions of pumpkin seed meal protein isolate (PSMPI) by response surface methodology.

Tasks to be solved in the research process:

1. Obtain a quadratic model of the extraction yield of pumpkin seed meal protein isolate (PSMPI).
2. Analyze how extraction factors, such as ultrasound power, ultrasound time, and the ratio of liquid to solid, affect the extraction yield of PSMPI by analyzing the response surface contour plots.
3. Experimentally determine the optimal conditions for the extraction of PSMPI.

**Methods.** In this study, defatted pumpkin seed meal and water were mixed in a 250 mL beaker. The pH of the mixture was adjusted and maintained to 10.5 by using a 2 N NaOH solution. The ultrasonic experiments were performed at 25 MHz using an Ultrasonic microwave synergistic extraction apparatus (XH-300B, Beijing xianghu, Beijing, China) with an ultrasonic unit (maximal power of 1500 W). On the basis of previous experiments, the extraction process was performed at different ultrasonic power ( $X_1$ : 400–500 W), ultrasonic time ( $X_2$ : 15–25 min) and ratio of liquid to solid ( $X_3$ : 20 to 30 mg/L) (Table 1).

After extraction, the product was centrifuged at  $4,000 \times g$  for 20 min, and the resulting supernatant was filtered to further remove the undissolved components. The filtrate was acidified to pH 4.5 with 2 N HCl, then centrifuged at  $4,000 \times g$  for 15 min. The pellet was recovered, washed with distilled water and centrifuged again as above. The pellet was freeze dried and used for protein determination and the yield calculation of PSMPI.

The experiment was conducted with triplicate samples. Analysis of variance was utilized to test for significant differences between independent variables ( $P < 0.05$ ). The statistical analysis was performed with Design expert 10 software (Stat-Ease, Minneapolis, MN, USA) and Microsoft Office Excel 2016 (Microsoft Corporation, USA), respectively.

**Results and discussion.** The independent variables  $X_1$  (ultrasonic power),  $X_2$  (ultrasonic time) and  $X_3$  (ratio of liquid to solid), at three variation levels studied were showed in Table 1.

Table 1

**Independent variables and levels for Box–Behnken design**

Independent variables	Symbol	Level		
		-1	0	1
Ultrasonic power (W)	$X_1$	400	450	500
Ultrasonic time (min)	$X_2$	15	20	25
Ratio of liquid to solid (mg/L)	$X_3$	20	25	30

The yield of pumpkin seed meal protein isolate (PSMPI) under the Box-Behnken experimental design was presented in Table 2. The quadratic model applied in the response surface was as follows:

$$Y = 80.93 + 1.63X_1 + 3.04X_2 + 1.27X_3 - 4.46X_1^2 - 3.38X_2^2 - 1.27X_3^2 - 0.042X_1X_2 - 1.52X_1X_3 - 0.74X_2X_3$$

According to the results of analysis of variance (ANOVA) (Table 3), the ultrasonic power, ultrasonic time, and ratio of liquid to solid all had significant positive linear effects. The coefficient of determination ( $R^2$ ) was 0.9681, which indicated a good model fit for 96.81% of the total variation can be explained by the model. The value of the adjusted determination coefficient ( $R_{Adj}^2$ ) was 0.9107, which also suggested that the model was significant. The values of coefficient variation (CV) was 1.61, indicating a very high degree of precision and a good reliability of the experimental values.

The effects of ultrasonic power ( $X_1$ ) and ultrasonic time ( $X_2$ ) on the extraction yield of PSMPI were pre-

sented in Fig. 1a, b, while ratio of liquid to solid was settled at its middle level (25 mg/L). As the response surface and its corresponding counter plot shows, the extraction yield of PSMPI increased slowly as the ultrasonic time increased when ultrasonic power at a low level. With the increase in ultrasonic power, the yield of PSMPI improved from about 75 g/100g to 80 g/100g at a high level of ultrasonic time. However, the yield of PSMPI increased only from about 71 g/100g to 75 g/100g at a low level of ultrasonic time. It indicated that the increase of ultrasonic power and ultrasonic time in the test range was beneficial for the extraction of PSMPI.

The response surface and its corresponding counter plot which showed the effects of ultrasonic power ( $X_1$ ) and ratio of liquid to solid ( $X_3$ ) on the yield of PSMPI were presented in Fig. 1c, d, while maintaining ultrasonic time ( $X_2$ ) at a middle level of 20 min. At the middle level of ultrasonic power and ratio of liquid to solid, the maximum extraction yield of PSMPI was achieved, further increasing in ultrasonic power and ratio of liquid to solid would not add into increase in the extraction yield of PSMPI.

The effects of ultrasonic time ( $X_2$ ) and ratio of liquid to solid ( $X_3$ ) on the yield of PSMPI were showed in Fig. 1e, f, while keeping the ultrasonic power ( $X_1$ ) at a middle level (450 W). The yield of PSMPI showed a considerable increase as the increase in ratio of liquid to solid at a high level of ultrasonic time. However, when ultrasonic time is at a low level,

Table 2

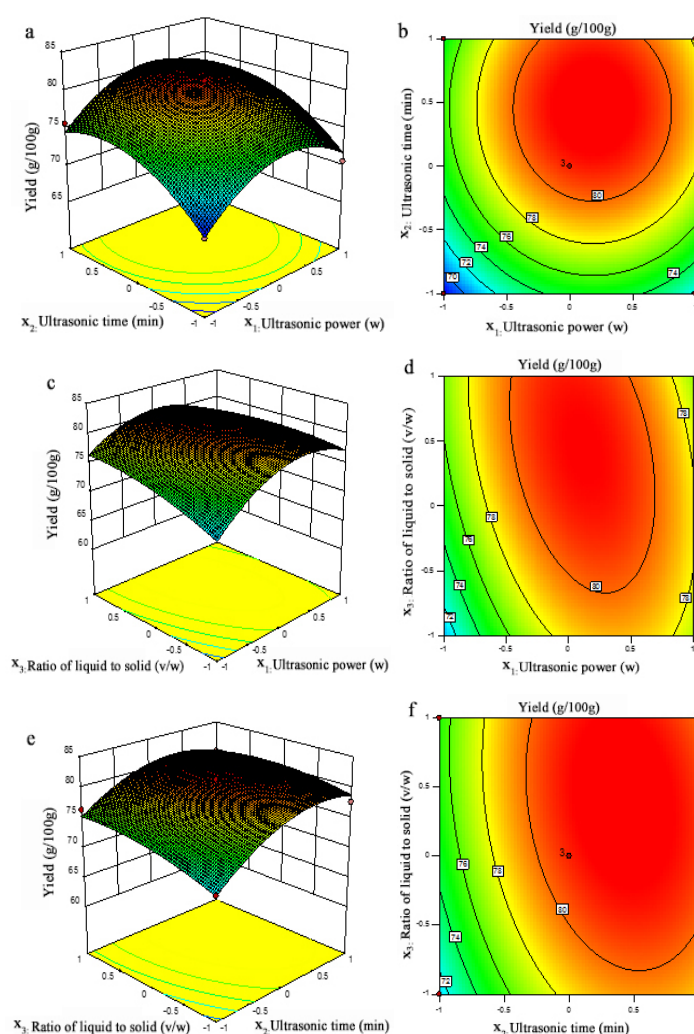
**Box-Behnken design arrangement, responses for protein yield of ultrasound-assisted alkaline extraction on pumpkin seed meal protein isolate (PSMPI)**

Run	$X_1$ : Ultrasonic power (W)	$X_2$ : Ultrasonic time (min)	$X_3$ : Ratio of liquid to solid (mg/L)	Yield (g/100 g)	Predicted Value (g/100 g)
1	1	1	0	77.79	77.72
2	1	0	-1	78.13	77.09
3	0	0	0	80.56	80.93
4	1	-1	0	70.59	71.72
5	1	0	1	76.59	76.57
6	-1	-1	0	68.31	68.38
7	0	1	1	79.77	79.86
8	-1	0	1	75.32	76.36
9	-1	0	-1	70.77	70.79
10	0	-1	-1	71.34	71.25
11	-1	1	0	75.68	74.55
12	0	1	-1	77.68	78.80
13	0	-1	1	76.37	75.25
14	0	0	0	80.95	80.93
15	0	0	0	81.29	80.93

the effect of ratio of liquid to solid on the yield of PSMPI was not significant.

The optimum extraction conditions depended on ultrasonic power ( $X_1$ ), ultrasonic time ( $X_2$ ) and ratio of liquid to solid ( $X_3$ ), and were obtained by response surface methodology (RSM). To validate the model, PSMPI were extracted under optimal UAEE conditions and its efficiency was determined. The result showed that the experimental value (80.76 g/100g) of the yield of PSMPI was reasonably close to the predicted one (81.86 g/100g). This suggested that there was a high fit degree between experimental value and the predicted value from the regression model. Hence, the response surface modeling can be utilized effectively to predict the extraction condition of the PSMPI.

**Conclusions.** In the present study, ultrasound-assisted alkaline extraction (UAEE) of pumpkin seed meal protein isolate (PSMPI) was proposed. Since many factors, like ultrasonic power, ultrasonic time and ratio of liquid to solid effect the yield of PSMPI, the response surface method was conducted to optimize the extraction conditions by three factors at three level Box-Behnken design. Response surface and contour plots suggested that the optimum condition for extracting PSMPI is at 456 W of ultrasonic power, 22 minutes of ultrasonic time and 27 mg/L for ratio of liquid to solid with a maximum protein yield of 81.86 g/100g. These results will contribute to the information of the productivity of the PSMPI by the method of UAEE.



**Fig. 1.** Response surfaces (a, c and e) and contour plots (b, d and f) showing effects of extraction variables ( $X_1$ , ultrasonic power;  $X_2$ , ultrasonic time;  $X_3$ , ratio of liquid to solid) on the ultrasound-assisted alkaline extraction (UAEE) yield of pumpkin seed meal protein isolate (PSMPI)

Table 3

**Analysis of the variance (ANOVA) of the quadratic model**

Source	Sum of Squares	df	Mean Square	F-Value	p-value Prob > F
Model	228.95	9	25.44	16.87	0.0031
$A_1$	21.19	1	21.19	14.05	0.0133
$A_2$	73.87	1	73.87	48.98	0.0009
$A_3$	12.83	1	12.83	8.50	0.0332
$A_{12}$	0.0072	1	0.0072	0.0048	0.9475
$A_{13}$	9.27	1	9.27	6.15	0.0559
$A_{23}$	2.16	1	2.16	1.43	0.2850
$A_{11}$	73.58	1	73.58	48.78	0.0009
$A_{22}$	42.10	1	42.10	27.91	0.0032
$A_{33}$	5.92	1	5.92	3.93	0.1043
Residual	7.54	5	1.51		
Lack of Fit	7.27	3	2.42	18.17	0.0526
Pure Error	0.27	2	0.13		
Cor Total	236.49	14			

$$R^2=0.9681 \quad R_{Adj}^2=0.9107 \quad CV=1.61$$



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#### Геліх А.О., Дан Гао, Шеньхуа Дуан. ОПТИМІЗАЦІЯ УЛЬТРАЗВУКОВОЇ ПІДТРИМКИ ЛУЖНИЙ ЕКСТРАКЦІЇ ІЗОЛЯТИВ БІЛКА БОРОШНА З ГАРБУЗОВОГО НАСІННЯ ЗА МЕТОДОЛОГІЄЮ ПОВЕРХНІ ВІДГУКУ

В якості альтернативи тваринним білкам ізолят білка борошна з насіння гарбуза (ІББНГ) може застосовуватися в харчовій промисловості в якості харчових добавок і функціональних харчових продуктів. Звичайний процес екстракції білка представляє собою лужний розчин і кислотні осадки, які перетворюють тривалий час. Ультразвуковий метод є екологічно чистим методом, який можна використовувати для підвищення ефективності екстракції білка. У цьому дослідженні був розроблений метод ультразвукової лужної екстракції (УЛЕ) білка борошна з насіння гарбуза. Методологія поверхні відгуку (МПВ) була використана для оптимізації умов УЛЕ (потужність ультразвуку, час ультразвуку і співвідношення рідини до твердого речовини) шляхом реалізації трьохфакторної трирівневої конструкції Боксу-Бенкена. Квадратичне поліноміальне рівняння було отримано за допомогою поліноміального регресійного аналізу експериментальних даних і відповідних статистичних методів. Вплив взаємодії незалежних змінних на вихід екстракції ІББНГ було проілюстровано за допомогою поверхонь відгуку та контрольних графіків. Крім того, шляхом вирішення рівняння регресії і аналізу контурних графіків поверхні відгуку були отримані певні оптимальні умови екстракції ІББНГ, які мали потужність ультразвуку 456 Вт, час ультразвуку 22 хв і співвідношення рідини до твердої речовини 27 мл / м. Була визначена ефективність УЛЕ по виходу ІББНГ. Результат показав, що експериментальне значення (80,76 г / 100 г) виходу ІББНГ було досить близько до прогнозованого (81,86 г / 100 г). Це говорить про те, що існує висока ступінь відповідності між експериментальним значенням і прогнозованим значенням з регресійної моделі. Таким чином, доведено, що рівняння поверхні відгуку може ефективно використовуватися для прогнозування умови вилучення ІББНГ.

**Ключові слова:** Бокс-Бенкена, ультразвукова лужна екстракція, білковий ізолят борошна з насіння гарбуза, оптимізована модель.